

MutaPLEX® STI-2 (TV/MH/UPU) real-time-PCR Kit

*For in vitro detection of the DNA of Trichomonas vaginalis,
Ureaplasma parvum, Ureaplasma urealyticum and
Mycoplasma hominis extracted from biological
specimens.*

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KG197296



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1 INTENDED PURPOSE

The MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit is designed for the qualitative detection of the nucleic acid of *Trichomonas vaginalis*, *Ureaplasma parvum*, *Ureaplasma urealyticum* and *Mycoplasma hominis* in eluates from biological specimens. The assay is an in vitro diagnostic medical device and intended to be used by professional users in a laboratory environment. It can be performed manually or using an automated platform. The assay serves as an aid in the diagnosis, screening and monitoring of *Trichomonas vaginalis*, *Ureaplasma parvum*, *Ureaplasma urealyticum* and *Mycoplasma hominis*.

2 PATHOGEN INFORMATION

Trichomonas vaginalis is an anaerobic, flagellated protozoan parasite. It is the causative agent of the sexual transmitted disease trichomoniasis. While infections in men are usually asymptomatic, infections in women usually comes along with symptoms of vaginitis [1] [2].

Ureaplasma parvum, *Ureaplasma urealyticum* and *Mycoplasma hominis* belong to the family of Mycoplasmatales. The Mycoplasmatales are small bacteria that lack a cell wall and possess a three-layered cellular membrane. Therefore, these bacteria are resistant to a lot of different antibiotics targeting the cell wall synthesis.

Both *Ureaplasma* species are known to cause inflammatory vulvovaginitis, urethritis, bacterial vaginosis and infertility. The ability to perform urea hydrolysis which creates ammonia as a product which comes along with a characteristic smell.

Mycoplasma hominis is associated with pelvic inflammatory disease, vaginal bacteriosis and male infertility [3].

3 PRINCIPLE OF THE TEST

The MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit contains specific primers and dual-labelled probes for the amplification of the DNA of *Trichomonas vaginalis*, *Ureaplasma parvum*, *Ureaplasma urealyticum* and *Mycoplasma hominis* extracted from biological specimens.

The presence of nucleic acid is detected by an increase in fluorescence due to hydrolysis of the probes during amplification. The fluorescence of the *Trichomonas vaginalis* specific probes is measured in the FAM channel. The fluorescence of the *Ureaplasma* specific probes is measured in the ROX channel. The fluorescence of the *Mycoplasma hominis* specific probes is measured in the Cy5 channel.

Furthermore, MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit contains a Control DNA (Internal Process Control, IPC), which is added during DNA extraction and detected in the same reaction by a HEX-labeled probe.

The Control DNA allows the detection of PCR inhibition and acts as control, that the nucleic acid was isolated from the biological specimen.

4 PACKAGE CONTENTS

The reagents supplied are sufficient for 96 reactions.

Table 1: Components of the MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit

Label	Lid Colour	Content
Reaction Mix	yellow	1 x 1344 µl
Positive Control	red	1 x 150 µl
Negative Control	green	1 x 150 µl
Control DNA	colourless	1 x 480 µl

5 EQUIPMENT AND REAGENTS TO BE SUPPLIED BY USER

- DNA isolation kit (e.g. MutaCLEAN® Mag RNA/DNA, KG1023 or KG1024).
- PCR grade water
- Sterile microtubes
- Calibrated precision pipets (adjustable volume) and sterile single-use tips with filter
- Table centrifuge
- Vortex
- Real time PCR instrument
- Optical PCR reaction tubes with lid or optical PCR reaction plate with optical foil
- Optional: Liquid handling system for automation

6 TRANSPORT, STORAGE AND STABILITY

The MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit is shipped on dry ice. All components must be stored at maximum -20°C in the dark immediately after receipt. Do not use reagents after the date of expiry printed on the package. Up to 20 freeze and thaw cycles are possible. Protect kit components from direct sunlight during the complete test run.

7 WARNINGS AND PRECAUTIONS

Read the Instruction for Use carefully before using the product.

Before first use check the product and its components for:

- Use of this product is limited to personnel specially instructed and trained in the techniques of Real-Time PCR procedures.
- Specimens should always be treated as infectious and/or biohazardous in accordance with safe laboratory procedures.
- Avoid microbial and nuclease (DNase/RNase) contamination of the eluates and the components of the kit.
- Always use DNase/RNase-free disposable pipette tips with aerosol barriers.
- Always wear protective disposable powder-free gloves when handling kit components.
- Use separated and segregated working areas for (1) sample preparation, (2) reaction setup and (3) amplification/detection activities. The workflow in the laboratory should proceed in unidirectional manner. Always wear disposable gloves in each area and change them before entering a different area.
- Dedicate supplies and equipment to the separate working areas and do not move them from one area to another.
- Store positive and/or potentially positive material separated from all other components of the kit.
- Do not open the reaction tubes/plates post amplification, to avoid contamination with amplicons.
- Additional controls may be tested according to guidelines or requirements of local, state and/or federal regulations or accrediting organisations.
- Do not autoclave reaction tubes after the PCR, since this will not degrade the amplified nucleic acid and will bear the risk to contaminate the laboratory area.
- Discard sample and assay waste according to your local safety regulations.
- Do not combine MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit components of different lot numbers.

8 SAMPLE MATERIAL

Starting material for MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit is DNA isolated from biological specimens. By the nature of the pathogens, sample material like vaginal swabs or urine are commonly used.

9 SAMPLE PREPARATION

Commercial kits for DNA isolation such as the following are recommended:

- MutaCLEAN® Mag RNA/DNA, Immundiagnostik Cat. No. KG1023 or KG1024

Please follow the Instructions for Use of the respective extraction kit.

Important:

In addition to the samples always run a ‚water control‘ in your extraction. Treat this water control analogous to a sample.

Comparing the amplification of the Control DNA in the samples to the amplification of the internal control in the water control will give insights on possible inhibitions of the real time PCR. Furthermore, possible contaminations during DNA extraction will be detectable.

Please note the chapter ‚Control DNA‘

If the real time PCR is not performed immediately, store extracted DNA according to the instructions given by the manufacturer.

10 CONTROL DNA

A Control DNA is supplied as extraction control. This allows the user to control the DNA isolation procedure and to check for possible real time PCR inhibition.

Add 5 µl Control DNA per extraction (5 µl x (N+1)). Mix well. Perform the DNA isolation according to the manufacturer's instructions.

The Control DNA must be added to the Lysis Buffer of the extraction kit.

11 REAL-TIME-PCR

11.1 Important Points Before Starting

- Please pay attention to the chapter 7 ‚Warnings and Precautions‘.
- Before setting up the real time PCR familiarise yourself with the real time PCR instrument and read the user manual supplied with the instrument.

- The programming of the thermal profile should take place before the PCR set up.
- In every PCR run one Positive Control and one Negative Control should be included.
- Before each use, all reagents should be thawed completely at room temperature, thoroughly mixed, and centrifuged very briefly.

11.2 Procedure

The Master Mix contains all of the components needed for the real time PCR except the sample. Prepare a volume of Master Mix for at least one sample more than required, in order to compensate for pipetting inaccuracy.

Table 2: Preparation of the master mix

Volume per reaction	Volume master mix
14.0 µl Reaction Mix	14.0 µl x (N+1)

11.3 Real time PCR set-up

- Place the number of optical PCR reaction tubes needed into the respective tray of the real time PCR instrument / take an optical PCR reaction plate.
- Pipet **14 µl** of the Master Mix into each optical PCR reaction tube / the optical PCR reaction plate.
- Add **6 µl** of the eluates from the DNA isolation (including the eluate of the water control), the Positive Control and the Negative Control to the corresponding optical PCR reaction tube / the optical PCR reaction plate (Table 3).
- Close the optical PCR reaction tubes / the optical PCR reaction plate immediately after filling in order to reduce the risk of contamination.

Table 3 Preparation of the real-time-PCR

Component	Volume
Master mix	14.0 µl
Sample	6.0 µl
Total volume	20.0 µl

11.4 Instrument Settings

For the real time PCR use the thermal profile shown in Table 4.

Table 4: real-time-PCR thermal profile

Description	Time	Temperature	Number of Cycles	Aquisition
Initial Denaturation	5 min	95 °C	1	no
<i>Amplification of DNA</i>				
Denaturation	10 sec	95 °C	45	no
Annealing and Extension	40 sec	60 °C		end of step

If in the same run samples should be tested for pathogens with RNA genome, use the thermal profile shown in Table 5.

Table 5: real-time-RT-PCR thermal profile

Description	Time	Temperature	Number of Cycles	Aquisition
Reverse Transcription	10 min	45 °C	1	no
Initial Denaturation	5 min	95 °C	1	no
<i>Amplification of DNA</i>				
Denaturation	10 sec	95 °C	45	no
Annealing and Extension	40 sec	60 °C		end of step

Dependent on the real time instrument used, further instrument settings have to be adjusted according to Table 6.

Table 6 Overview of the instrument settings required for the MutaPLEX® STI-2 (TV/MH/UPU) real time PCR.

Real time PCR instrument	Parameter	Detection channel	Notes		
LightCycler 480II	<i>T. vaginalis</i> Control DNA (IPC) <i>U. urealyticum, U. parvum</i> <i>M. hominis</i>	465–510 533–580 533–610 618–660	Colour Compensation Kit MutaPLEX® CC-1 (KG19-5-CC) required		
			Melt factor	Quant factor	Max integration time (s)
			1	10	1
			1	10	2
			1	10	2
1	10	3			
Stratagene Mx3000P/ Mx3005P	<i>T. vaginalis</i> Control DNA (IPC) <i>U. urealyticum, U. parvum</i> <i>M. hominis</i>	FAM HEX ROX Cy5	Gain 8 Gain 1 Gain 1 Gain 4	Reference Dye: None	
QuantStudio 5 CFX96 CFX96 Opus Aria Mx qTower ³ G	<i>T. vaginalis</i> Control DNA (IPC) <i>U. urealyticum, U. parvum</i> <i>M. hominis</i>	FAM HEX ROX Cy5	Option Reference Dye ROX: NO		
Mic qPCR Cycler	<i>T. vaginalis</i> Control DNA (IPC) <i>U. urealyticum, U. parvum</i> <i>M. hominis</i>	Green Yellow Orange Red	Gain 8 Gain 10 Gain 10 Gain 10		

12 DATA ANALYSIS

Following results can occur:

Signal / C _T Values				Interpretation
FAM Channel	Cy5 Channel	ROX Channel	HEX Channel	
<i>Trichomonas vaginalis</i>	<i>Mycoplasma hominis</i>	<i>U. parvum/ U. urealyticum</i>	Control DNA (IPC)	
positive	negative	negative	positive or negative ¹	Positive result. The sample contains <i>Trichomonas vaginalis</i> DNA.
negative	positive	negative	positive or negative ¹	Positive result. The sample contains <i>Mycoplasma hominis</i> DNA.
negative	negative	positive	positive or negative ¹	Positive result. The sample contains <i>Ureaplasma parvum</i> or <i>Ureaplasma urealyticum</i> DNA.
negative	negative	negative	≤ 34 ²	Negative result. The sample contains no <i>Trichomonas vaginalis</i> , <i>Mycoplasma hominis</i> , <i>Ureaplasma parvum</i> or <i>Ureaplasma urealyticum</i> DNA.
negative	negative	negative	negative or > 34 ²	No diagnostic statement can be made. The real time PCR is either inhibited or errors occurred while DNA extraction.

¹ A strong positive signal in the FAM, ROX or the Cy5 can inhibit the IPC. In such cases the result for the Control DNA can be neglected.

² In case of high C_T values, the IPC should be compared to the water extraction control as described in the chapter 'Assay validation'.

13 ASSAY VALIDATION

Negative Control

The Negative Control must show no C_T in the FAM, Cy5, ROX and HEX channel.

Positive control

The Positive Control must show a positive (i.e. exponential) amplification curve in the different channels FAM, Cy5 and ROX. The Positive Control must fall below C_T 30.

Internal Controls

The following values for the amplification of the internal controls are valid using Im-mundiagnostik nucleic acid extraction kit MutaCLEAN® Mag RNA/DNA. The Control DNA (IPC) must show a positive (i.e. exponential) amplification curve.

The IPC must fall below a C_T of 34. If the IPC is above C_T 34 this points to a purification problem or a strong positive sample that can inhibit the IPC. In the latter case, the assay is valid. It is recommended to perform the extraction of a water control in each run. The IPC in the water control must fall below a C_T of 34.

If other nucleic acid extraction kits are used, the customer must define own cutoffs. In this case the C_T value of the Control DNA (IPC) in an eluate from a sample should not be delayed for more than 4 C_T in comparison to an eluate from an extracted water control.

14 LIMITATIONS OF THE METHOD

- Strict compliance with the instructions for use is required for optimal results.
- Use of this product is limited to personnel specially instructed and trained in the techniques of real-time PCR and *in vitro* diagnostic procedures.
- Good laboratory practice is essential for proper performance of this assay.
- All reagents should be closely monitored for impurity and contamination. Any suspicious reagents should be discarded.
- This assay must not be used on a biological specimen directly. Appropriate nucleic acid extraction methods have to be conducted prior to using this assay.
- The presence of PCR inhibitors may cause false negative or invalid results.
- As with any diagnostic test, results of the MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit need to be interpreted in consideration of all clinical and laboratory findings.

15 TROUBLESHOOTING

The following troubleshooting guide is included to help you with possible problems that may arise when performing a real-time PCR. If you have further questions, please do not hesitate to contact our scientists on info@immundiagnostik.com.

No fluorescence signal in the FAM, Cy5, ROX channel of the Positive Control.

The selected channel for analysis does not comply with the protocol

Select the FAM channel for analysis of the *T. vaginalis* specific amplification, the Cy5 channel for the *M. hominis* specific amplification, the ROX channel for the amplification of *U. parvum* and *U. urealyticum* and the HEX channel for the amplification of the Control DNA (IPC).

Incorrect configuration of the real-time-PCR

Check your work steps and compare with chapter 11.

The programming of the thermal profile is incorrect

Compare the thermal profile with the protocol 'in chapter 11.4.

Incorrect storage conditions for one or more kit components or kit expired

Check the storage conditions and the date of expiry printed on the kit label. If necessary, use a new kit and make sure kit components are stored as described in 'Transport, storage and stability' (chapter 6).

Weak or no signal of the Control DNA (IPC) and simultaneous absence of a signal in the specific FAM and/or Cy5 and/or ROX channel.

real time PCR conditions do not comply with the protocol

Check the real time PCR conditions in Table 4.

real time PCR inhibited

Make sure that you use an appropriate isolation method (see chapter 'Sample Preparation') and follow the manufacturer's instructions. Make sure that the ethanol-containing washing buffers have been completely removed.

sample material not sufficient

Make sure that enough sample material has been applied to the extraction. Use an appropriate isolation method (see chapter 9 - 'Sample Preparation') and follow the manufacturer's instructions.

DNA loss during isolation process

Lack of an amplification signal in the HEX channel can indicate that the DNA isolation was not successful. Make sure that you use an appropriate isolation method (commercial kits are recommended) and stick to the manufacturer's protocol.

Incorrect storage conditions for one or more components or kit expired

Check the storage conditions and the date of expiry printed on the kit label. If necessary, use a new kit and make sure kit components are stored as described in 'Transport, Storage and Stability (chapter 6).

Detection of a fluorescence signal in the FAM and/or ROX and/or Cy5 channel of the Negative Control.***Contamination during preparation of the real-time PCR***

Repeat the real time PCR in replicates. If the result is negative in the repetition, the contamination occurred when the samples were pipetted into the optical PCR reaction tubes. Make sure to pipet the Positive Control last and close the optical PCR reaction tube immediately after adding the sample. If the same result occurs, one or more of the kit components might be contaminated. Make sure that work space and instruments are decontaminated regularly. Use a new kit and repeat the real time PCR.

16 KIT PERFORMANCE***16.1 Analytical Sensitivity***

For the FAM, ROX and Cy5 channels, the limits of detection (LoD) of MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit were determined using serial dilutions of the of synthetic DNA-fragments containing the specific gene target sequence. The determination of the LoD was done on a CFX Opus 96 Instrument (Bio-Rad).

The LoD of MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit is ≤ 2.5 genome copies per μl for the FAM, Cy5 and ROX channel.

16.2 Analytical Specificity

The specificity of the MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit was evaluated with different ring trial samples of known status and different other relevant viruses and bacteria found in biological samples and basing on in silico analyses. Additionally, 321 qualified field samples were tested with the MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit.

All ring trial samples and other eluates with known status were detected correctly. Results are shown in Table 7 and Table 8.

The results for the field samples are summarized for each target in Table 9, Table 10 and Table 11.

Table 7: Ring trial samples tested for the validation of the sensitivity and specificity of the MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit.

sample	<i>T. vaginalis</i>	<i>M. hominis</i>	<i>U. parvum / U. urealyticum</i>
	FAM channel	Cy5 channel	ROX channel
QCMD 2020 Sexually Transmitted Infections I			
STI_I101S-01 Trichomonas vaginalis	positive	negative	negative
STI_I101S-02 Mycoplasma hominis	negative	positive	negative
STI_I101S-03 G. vaginalis + T. vaginalis	positive	negative	negative
STI_I101S-04 M. genitalium (drug resistant)	negative	negative	negative
STI_I101S-05 M. genitalium (wild type)	negative	negative	negative
STI_I101S-06 negative	negative	negative	negative
STI_I101S-07 Gardnerella vaginalis	negative	negative	negative
STI_I101S-08 Trichomonas vaginalis	positive	negative	negative
STI_I101S-09 M. hominis + C. trachomatis	negative	positive	negative
STI_I101S-10 Trichomonas vaginalis	positive	negative	negative
QCMD 2020 Sexually Transmitted Infections II			
STI_II101S-01 Herpes simplex virus (type 2)	negative	negative	negative
STI_II101S-02 Treponema pallidum	negative	negative	negative
STI_II101S-03 Herpes simplex virus (type 1)	negative	negative	negative
STI_II101S-04 C. trachomatis + M. hominis	negative	positive	negative
STI_II101S-05 Neisseria gonorrhoeae	negative	negative	negative
STI_II101S-06 C. trachomatis + N. gonorrhoeae + M. hominis	negative	positive	negative
STI_II101S-07 C. trachomatis + M. hominis	negative	positive	negative
STI_II101S-08 Neisseria gonorrhoeae	negative	negative	negative
STI_II101S-09 Chlamydia trachomatis	negative	negative	negative
STI_II101S-10 Neisseria gonorrhoeae	negative	negative	negative
QCMD 2020 Trichomonas vaginalis			
TV101S-01 Trichomonas vaginalis	positive	negative	negative

sample	<i>T. vaginalis</i>	<i>M. hominis</i>	<i>U. parvum / U. urealyticum</i>
	FAM channel	Cy5 channel	ROX channel
TV101S-02 Trichomonas vaginalis	positive	negative	negative
TV101S-03 T. vaginalis + C. trachomatis + M. hominis	positive	positive	negative
TV101S-04 T. vaginalis + G. vaginalis	positive	negative	negative
TV101S-05 Trichomonas negative	negative	negative	negative
TV101S-06 Trichomonas vaginalis	positive	negative	negative
TV101S-07 Trichomonas vaginalis	positive	negative	negative

Table 8: Eluted DNA/RNA from bacterial and viral pathogens tested for the determination of the analytical specificity of MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit.

Eluates with known status	MutaPLEX® STI-2 (TV/MH/UPU)		
	<i>T. vaginalis</i>	<i>M. hominis</i>	<i>U. parvum / U. urealyticum</i>
	FAM channel	Cy5 channel	ROX channel
Cytomegalievirus	negative	negative	negative
Chlamydia pneumoniae	negative	negative	negative
Herpes Simplex Virus Type 1	negative	negative	negative
Herpes Simplex Virus Type 2	negative	negative	negative
Mycoplasma pneumoniae	negative	negative	negative
Mycoplasma hyopneumoniae	negative	negative	negative
Treponema phagadenis	negative	negative	negative
Varizella Zoster Virus Genotype 3	negative	negative	negative
Varizella Zoster Virus Genotype 5	negative	negative	negative
Chlamydia trachomatis	negative	negative	negative
Mycoplasma genitalium	negative	negative	negative
Neisseria gonorrhoeae	negative	negative	negative
Gardnerella vaginalis	negative	negative	negative
Trichomonas vaginalis	positive	negative	negative
Mycoplasma hominis	negative	positive	negative
Ureaplasma parvum	negative	negative	positive
Ureaplasma urealyticum	negative	negative	positive

Table 9: Qualified field samples tested for *T. vaginalis* using MutaPLEX® STI-2 (TV/MH/UPU) real time PCR.

		<i>Trichomonas vaginalis</i>	
		positive	negative
MutaPLEX® STI-2 (TV/MH/UPU) (KG197296)	positive	2	0
	negative	0	319
		Sensitivity (%)	Specificity (%)
		100	100

Table 10: Qualified field samples tested for *M. hominis* using MutaPLEX® STI-2 (TV/MH/UPU) real time PCR.

		<i>Mycoplasma hominis</i>	
		positive	negative
MutaPLEX® STI-2 (TV/MH/UPU) (KG197296)	positive	26	0
	negative	0	295
		Sensitivity (%)	Specificity (%)
		100	100

Table 11: Qualified field samples tested for *U. parvum* and *U. urealyticum* using MutaPLEX® STI-2 (TV/MH/UPU) real time PCR.

		<i>U. parvum</i> / <i>U. urealyticum</i>	
		positive	negative
MutaPLEX® STI-2 (TV/MH/UPU) (KG197296)	positive	140	0
	negative	0	181
		Sensitivity (%)	Specificity (%)
		100	100

16.3 Linear Range

The linear range of the MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit was evaluated by analysing logarithmic dilution series of quantified synthetic DNAs of the target sequences.

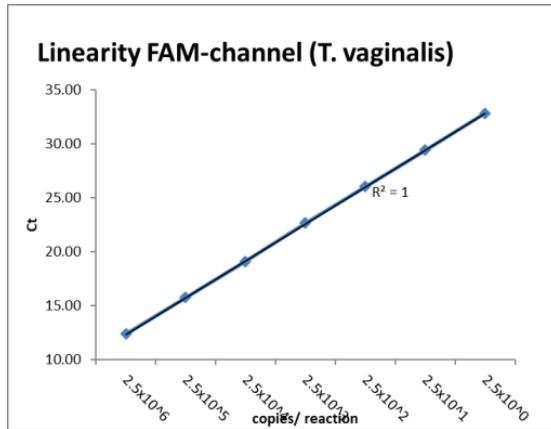


Figure 1: Determination of the linear range of MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit for *T. vaginalis* in the FAM channel.

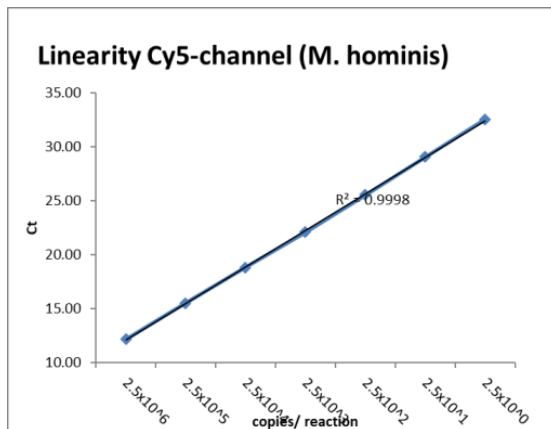


Figure 2: Determination of the linear range of MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit for *M. hominis* in the Cy5 channel.

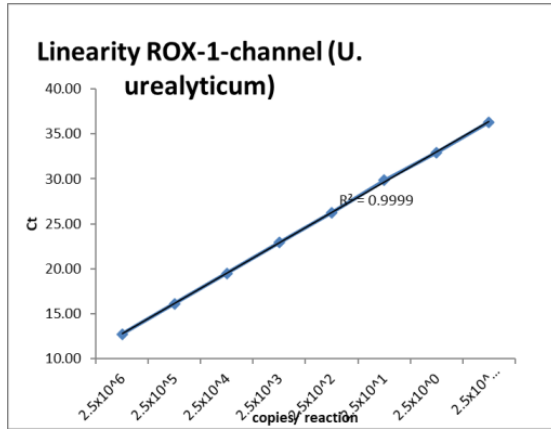


Figure 3: Determination of the linear range of MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit for *U. urealyticum* in the ROX channel

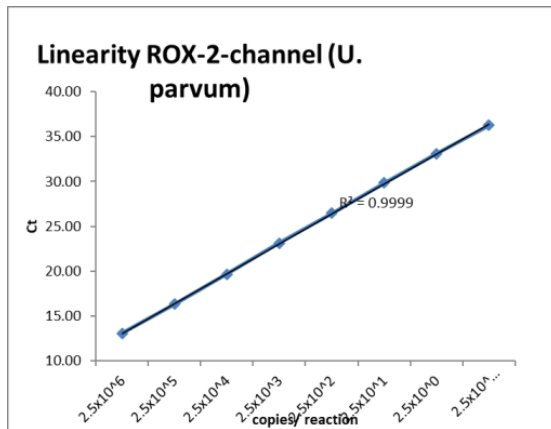


Figure 4: Determination of the linear range of MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit for *U. parvum* in the ROX channel.

16.4 Precision

The precision of the MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit was determined as intra-assay variability, inter-assay variability and inter-lot variability.

Variability data are expressed by standard deviation and coefficient of variation. The data are based on quantification analyses of defined concentrations of *T. vaginalis* specific synthetic DNA, *M. hominis* specific synthetic DNA, *U. parvum* and *U. urealyti-*

cum specific synthetic DNA and on the threshold cycle of the Control DNA (IPC). The results are shown in Table 12.

Table 12: Precision of the MutaPLEX® STI-2 (TV/MH/UPU) real time PCR Kit.

<i>T. vaginalis</i> (FAM)	copies/μl	Standard Deviation	Coefficient of Variation [%]
Intra-Assay Variability	2.5	0.37	1.13
Inter-Assay-Variability	2.5	0.61	1.85
Inter-Lot-Variability	2.5	0.28	0.86
<i>M. hominis</i> (Cy5)	copies/μl	Standard Deviation	Coefficient of Variation [%]
Intra-Assay Variability	2.5	0.42	1.30
Inter-Assay-Variability	2.5	0.61	1.88
Inter-Lot-Variability	2.5	0.07	0.23
<i>U. urealyticum</i> (ROX)	copies/μl	Standard Deviation	Coefficient of Variation [%]
Intra-Assay Variability	2.5	0.25	0.76
Inter-Assay-Variability	2.5	0.27	0.84
Inter-Lot-Variability	2.5	0.21	0.65
<i>U. parvum</i> (ROX)	copies/μl	Standard Deviation	Coefficient of Variation [%]
Intra-Assay Variability	2.5	0.33	1.00
Inter-Assay-Variability	2.5	0.53	1.61
Inter-Lot-Variability	2.5	0.17	0.52
IPC (HEX)	copies/μl	Standard Deviation	Coefficient of Variation [%]
Intra-Assay Variability	5000	0.17	0.58
Inter-Assay-Variability	5000	0.22	0.75
Inter-Lot-Variability	5000	0.27	0.90

16.5 *Diagnostic Sensitivity*









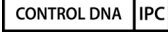







The diagnostic sensitivity of real time (RT-)PCR assays is mainly dependent on the DNA/RNA extraction method used to isolate DNA and RNA from various biological specimens. DNA/RNA extraction reagents are not part of the Immundiagnostik real time (RT-)PCR kits. Immundiagnostik real time (RT-)PCR kits include an extraction control and guidelines for the validation criteria of the extraction control in each reaction. The extraction control indicates inhibition of the real time (RT-)PCR and/or inefficient nucleic acid extraction. It cannot be used as a calibrator.

Therefore, Immundiagnostik guarantees the analytical sensitivities and specificities of the real time (RT-)PCR kits, performed with eluted DNA and RNA from reference materials and ring trial samples and with synthetic nucleic acid fragments. Immundiagnostik does not guarantee diagnostic sensitivities. If diagnostic sensitivities are mentioned in manuals of Immundiagnostik real time (RT-)PCR kits, the data are strictly correlated to a specific nucleic acid extraction method that has been used during the validation of the respective kits and cannot be transferred to other extraction methods. It is the responsibility of the user to qualify the extraction methods used for DNA/RNA isolation from biological samples.

17 LITERATURE

- [1] Herath et al. (2021) Comparison of diagnostic methods and analysis of socio-demographic factors associated with *Trichomonas vaginalis* infection in Sri Lanka. Plos One 16(10)
- [2] Frolund et al. (2014) Comparison between Culture and Multiplex Quantitative Real-Time Polymerase Chain Reaction Assay Detecting *Ureaplasma urealyticum* and *U. parvum*. Plos One 9(7)
- [3] Campos et al. (2015) Prevalence of *Mycoplasma genitalium* and *Mycoplasma hominis* in urogenital tract of Brazilian women. BMC Infectious Disease 15:60

18 ABBREVIATIONS AND SYMBOLS

DNA	(complementary) Deoxyribonucleic acid		Catalog number
PCR	Polymerase chain reaction		To be used with
	Reaction mix		Contains sufficient for <n> test
	Positive Control		Upper limit of temperature
	Negative Control		Manufacturer
	Control DNA (IPC)		Use by
	Unique Device Identification		Lot number
	<i>In vitro</i> diagnostic medical device		Content
	European conformity		Consult instructions for use

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